1	CIPOLLINI AND SCHRADIN: IMPACTS OF RANUNCULUS FICARIA
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3	Guilty in the Court of Public Opinion: Testing Presumptive Impacts and Allelopathic
4	Potential of Ranunculus ficaria
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9	ABSTRACT. –Information about invasive species is often based primarily on anecdotal
10	evidence, indicating the need for further information. Ranunculus ficaria is an ephemeral
11	riparian plant species that is presumed invasive in the United States, despite the lack of any
12	published information on its impacts. Mechanisms by which R. ficaria may affect native plant
13	species include competition and allelopathy. We examined if <i>R. ficaria</i> negatively affected the
14	growth and reproduction of the native Impatiens capensis and, if so, whether it is by allelopathy,
15	nutrient competition or some combination thereof. We performed a fully-factorial field
16	experiment, manipulating the presence of R. ficaria, nutrients, and allelopathy with the use of
17	activated carbon. The presence of R. ficaria tended to negatively affect life span of I. capensis. In
18	the absence of carbon, R. ficaria significantly decreased seed production, illustrating the negative
19	impact of R. ficaria. In the presence of carbon, there was no effect of R. ficaria, suggesting that
20	carbon may have ameliorated the negative allelopathic effect of R. ficaria. Nutrient competition
21	did not show strong effects. Despite its widespread identification as an invasive species, this is
22	the first study to demonstrate the negative impact of <i>R</i> . <i>ficaria</i> on a native species and the

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23	possible role of allelopathy in its success. Further, the negative impacts of this ephemeral species
24	persist well beyond its early growing season, which calls into question previous widespread
25	assumptions about R. ficaria exerting effects primarily on other ephemeral species.
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28	INTRODUCTION
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30	Invasive species threaten biodiversity on a global scale (Wilcove et al., 1998; McGeoch et al.,
31	2010) and are defined as those non-native species that cause or have the potential to cause
32	economic or environmental harm, weighed against their benefits (NISC, 2006). Most naturalized
33	plants are introduced through the horticultural industry (Mack and Erneberg, 2002). Some
34	horticultural species can still be purchased in some instances despite their invasive status (Harris
35	et al., 2009; Axtell et al., 2010). However, only a portion of naturalized species are actually
36	considered to be invasive (Milbau and Stout, 2008). There is much interest in characterizing the
37	species traits that make a species invasive and the routes by which they tend to be introduced
38	(Lambdon et al., 2008; Milbau and Stout, 2008; van Kleunen et al., 2010). Yet, in many of these
39	studies the methods by which species are even categorized as invasive are vague and based on
40	expert opinion and anecdotal evidence, with little scientific evidence (Blossey, 1999). Further,
41	even when there is some published information, the impacts of an invasive species can be
42	overstated by the popular press (Lavoie, 2010), which may lead to inappropriate response
43	strategies or undue focus. The lack of information on the impact of invasive species and on the
44	possible mechanism of impact is an obstacle to effectively prioritizing the control of invasive
45	species during a time of dwindling resources.

46 Invasive plant species can negatively impact native species through a variety of mechanisms 47 (Levine et al., 2003). Invasive species can simply outcompete native species for above- and/or 48 below-ground resources (e.g., Kueffer et al., 2007; Cipollini et al., 2008a; Galbraith-Kent and 49 Handel, 2008). Enhanced nutrient acquisition can lead to invasive species success. For example, 50 *Centaurea maculosa* acquired more phosphorus than surrounding native species which may have 51 increased competitive success (Thorpe et al., 2006). Additionally, Leishman and Thomson 52 (2005) found that in a study testing 28 different invasive species, the 15 invasive species had 53 greater responses on average to high nutrient soils than the 13 non-invasive species tested, thus 54 providing a possible mechanism for why invasive species may be able to outcompete natives in 55 nutrient-rich environments. Another mechanism by which invasive species affect native 56 communities is allelopathy (Hierro and Callaway, 2003; Ens et al. 2009). Most plants produce 57 secondary compounds (Ehrenfeld, 2006) that can affect an adjacent plant either directly 58 (Cipollini *et al.*, 2008b) or indirectly through changing soil ecology (Stinson *et al.*, 2006; Mangla 59 et al., 2008). Some allelopathic chemicals that have no negative impact in their native 60 environment may have negative effects in an invaded community, a mechanism coined the 61 "novel weapons hypothesis" (Callaway and Aschehoug, 2000; Callaway and Ridenour, 2004; 62 Callaway et al., 2008; Thorpe et al., 2009). 63 Discovering impacts due to allelopathy can be done with careful experimentation (Inderjit and Callaway, 2003). Allelopathic effects have been studied using activated carbon (e.g., Ridenour 64

and Callaway, 2001; Cipollini et al., 2008a). Activated carbon adsorbs organic compounds,

66 including allelochemicals (Ridenour and Callaway, 2001). Addition of carbon can also has

67 effects on soil properties and plant growth in potting soil (Lau *et al.*, 2008; Weisshuhn and Prati,

68 2009). Addition of nutrients is thought to help ameliorate any fertilizing effects of the addition of

69	activated carbon (Inderjit and Callaway, 2003). Activated carbon may also serve as a restoration
70	tool to change soil conditions in invaded soils (Kulmatiski and Beard, 2006), by a combination
71	of absorption of allelochemicals and/or effects on microbial communities (Kulmatiski, 2010).
72	Ranunculus ficaria L. (Ranunculaceae), or lesser celandine, is a groundcover native to Europe
73	(Taylor and Markham, 1978; Sell, 1994), which appears to be negatively affecting native plants
74	in forested floodplains in many US states (Swearingen, 2005). There are five known subspecies,
75	all of which are found in the United States (Post et al., 2009). Ranunculus ficaria was first
76	recorded naturalized in the United States in 1867 (Axtell et al., 2010). As it is still being
77	marketed by the nursery industry (Axtell et al., 2010), it was likely imported for horticultural
78	purposes. It was recognized as a naturalized species in the Midwest in the 1980's (Rabeler and
79	Crowder, 1985) and in southern states more recently (Krings et al., 2005; Nesom, 2008).
80	Ranunculus ficaria is currently documented in at least 21 US states, the District of Columbia,
81	and 4 Canadian provinces (USDA, 2010). It has been identified as invasive in 9 states and the
82	District of Columbia and is banned in Massachusetts and Connecticut as a noxious weed (Axtel
83	<i>et al.</i> , 2010).
84	Ranunculus ficaria emerges before most native spring species, which may provide it with a
85	competitive advantage. Once established, it spreads rapidly across the forest floor to form a
86	dense monoculture, which native species seemingly cannot penetrate (Swearingen, 2005).

87 Hammerschlag *et al.* (2002) reported that *R. ficaria* created a monoculture in the Rock Creek

88 floodplain in Washington, DC and few native species re-colonized after its removal. It is thought

89 that *R. ficaria*, as a spring ephemeral, has impacts primarily on other spring ephemerals

90 (Swearingen, 2005). However, most all information on *R. ficaria* as an invasive species is

91 primarily anecdotal in nature. Unpublished and preliminary data indicate that presence of *R*.

92	ficaria is associated with reduced abundance and richness of native species in spring and summer				
93	field surveys (Hohman, 2005). Ranunculus ficaria exhibits direct allelopathic effects on				
94	germination of some native species (Cipollini, unpublished data), indicating that R. ficaria may				
95	have negative effects beyond the spring time period through lingering allelopathic effects.				
96	For our study, we examined if <i>R. ficaria</i> negatively affects the growth and reproduction of the				
97	native annual Impatiens capensis and, if so, whether it is by allelopathy, nutrient competition or				
98	some combination thereof. In the field we performed a fully-factorial experiment with the				
99	treatments of R. ficaria (present and absent), carbon (present and absent), and nutrient addition				
100	(present and absent). We expected that the presence of <i>R. ficaria</i> would have an overall negative				
101	impact, that addition of nutrients would have an overall positive impact and that addition of				
102	carbon would have no overall effect on the performance of <i>I. capensis</i> . If allelopathy were				
103	important, we expected to see a significant carbon by <i>R</i> . <i>ficaria</i> interaction and, if nutrient				
104	competition were important, we expected to see a significant nutrient by R. ficaria interaction on				
105	I. capensis response variables.				
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108	METHODS				
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110	We performed the study in 2009 at Hamilton County Park District's Winton Woods in				
111	Cincinnati, Ohio in an area invaded by R. ficaria ssp. bulbilifer, a subspecies that forms asexual				
112	bulbils (Post et al., 2009). The study area was found in a floodplain along Westfork Mill Creek.				
113	We set up the experiment on 24 April, choosing an area with uniform 95-100% coverage of <i>R</i> .				
114	ficaria. We fenced the entire 3 x 4-m site using deer fencing to prevent trampling and/or				

115 herbivory. We used a fully factorial design with the main factors of: presence/absence of R. 116 *ficaria*, presence/absence of activated carbon and presence/absence of additional nutrients, 117 replicated 8 times (2 R. ficaria levels x 2 carbon levels x 2 nutrient levels x 8 replicates = 64 experimental units). The treatment combinations were haphazardly assigned to each 25 cm^2 plot 118 119 and each plot was located approximately 25 cm apart. Treatments were planted in 5 rows and 120 center rows could be accessed (with minimal disturbance to other plots) from the outside edge of 121 the experimental area. We tested the effects of these treatment combinations on the target plant 122 Impatiens capensis Meerb., jewelweed (Balsaminaceae). A single plant was used in each 123 experimental plot. We chose *I. capensis* because of the overlap in habitat and distribution with *R*. 124 *ficaria*. Additionally, the nearly complete lack of *I. capensis* within the *R. ficaria* monoculture at 125 our site indicated the potential negative impact of *R. ficaria* on *I. capensis*. Another advantage is 126 that reproduction can be measured in this annual species. Furthermore, *I. capensis* has served as 127 a model organism in previous studies of invasive species effects (e.g., Cipollini et al., 2008a; 128 Cipollini et al., 2009; Cipollini and Hurley, 2009). Any competitive effect or plot interaction 129 would be negligible at 25 cm. This distance between plants has been used in previous studies 130 (Cipollini et al., 2008a, Cipollini et al., 2009) and we have observed in this study and earlier 131 research that above-ground competition has been absent at this distance. Another study found 132 that the density-dependent effect on resistance to disease was not seen when plants were thinned to 15 cm apart (Lively et al., 1995), suggesting negative competitive effects are minimal at this 133 134 distance. Impatiens capensis usually grows in much higher densities; natural populations of I. capensis can be found at densities of more than 2,500 seedlings/m² (Schmitt et al., 1987). All I. 135 136 capensis seedlings were obtained in an immediately adjacent area free of R. ficaria.

137 In the *R. ficaria*-present treatments, we removed *R. ficaria* and planted them back in place (at 138 their natural density) while transplanting *I. capensis* seedlings. Identifying how many plants of *R*. 139 *ficaria* were back-transplanted in a non-destructive manner was nearly impossible in the dense 140 monoculture of clonally-reproducing R. ficaria. However, the back-transplantation of R. ficaria 141 was completely successful and led to nearly 100% R. ficaria cover in experimental plots with the 142 R. ficaria-present treatment. In R. ficaria-absent treatments, we removed the R. ficaria 143 completely in the 25 cm² plot before transplanting *I. capensis*. In activated carbon-present 144 treatments, we worked 10 ml of activated carbon (Aquarium Pharmaceuticals Black Magic 145 Activated Carbon, Chalfont, Pennsylvania) into the top 8 cm of soil of each plot. This ratio of 146 activated carbon to soil volume has been shown to mitigate allelopathic effects in previous 147 research (Ridenour and Callaway, 2001; Cipollini et al., 2008a). In nutrient-addition treatments, 148 we worked the manufacturer-recommended amount of 1.5 teaspoons of slow-release fertilizer 149 (Scotts Osmocote, Scotts-Sierra Horticultural Product Co., Marysville, Ohio) into the top 8 cm of 150 soil in each plot. We disturbed the soil in each plot in the same manner (to 8 cm deep) in every 151 plot regardless of treatment to control for any soil disturbance effects. We noticed no movement 152 of the large-sized carbon or fertilizer granules out of the experimental plots. No flooding 153 occurred during the course of the experiment. On May 1 we replaced any *I. capensis* transplants 154 that did not survive, presumably due to transplant shock. We measured the height of each 155 seedling when transplanted. The number of fruits, number of seeds and life span (days to death) 156 of the seedlings were recorded once each week. Height and stem diameter (measured directly 157 beneath bottom node with a digital caliper) were measured. Ranunculus ficaria had lost all of its 158 foliage by 2 June (week 6) and the leaf litter had decomposed by 12 June (week 8), leaving the

bulbils exposed on the soil surface. Measurements began on 5 May and ended on 28 August(week 18).

161 We performed separate Analysis of Variances (ANOVAs) on the *I. capensis* response 162 variables of life span and total number of seeds, using the full model with fixed factors of 163 nutrients (+/-), R. ficaria (+/-) and carbon (+/-). We used a General Linear Model (GLM), which 164 can be used for unbalanced designs, using the appropriate Type III sums of squares (Ryan et al., 165 2005). We were unable to include all of the variables in a single multivariate model due to the 166 missing values generated as plants died. For life span, we analyzed the day to death for all plants. 167 For total number of seeds, we removed the ten plants that had died within eight weeks of 168 transplant, as seed production was essentially non-existent up to that time. For final height and 169 stem diameter, we used a MANCOVA (using the full model as with the ANOVAs) with starting 170 height as a significant covariate for the 32 plants that survived to the last growth measurement on 171 August 14 (week 16) (SAS 1999; Scheiner 2001). When significance was found in the 172 MANCOVA using Wilk's λ , we ran separate univariate Analyses of Covariance (ANCOVAs) 173 for each variable. For all statistical tests, model assumptions of normality and heteroskedasticity 174 were verified prior to analysis and transformed where appropriate. 175 176 177 RESULTS 178 179 For life span, there was a near-significant effect of *R. ficaria* ($F_{1,55} = 3.75$, P = 0.058), with *I.* 180 capensis tending to die sooner when R. ficaria was present (Fig. 1). For the total number of seeds 181 produced, there was a significant effect of nutrients and a significant interactive effect between

182	R. ficaria-presence and carbon (Table 1). The presence of nutrients nearly tripled seed
183	production (Fig. 2). When carbon was absent, the presence of <i>R</i> . <i>ficaria</i> reduced seed production.
184	When carbon was present, seed production was similar whether R. ficaria was present or absent
185	(Fig. 3). In the MANCOVA, there was a significant effect of nutrients on final height and stem
186	diameter (Table 2; eigenvalue = 0.4002, canonical coefficient for height = 1.126 and diameter =
187	1.96). In the univariate ANCOVA, nutrients increased both height (Table 3) and stem diameter
188	(Table 4) (Fig. 2).
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191	DISCUSSION
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193	We wanted to know if the putative invasive <i>R</i> . <i>ficaria</i> negatively affected the growth and
194	reproduction of the native I. capensis, and if it did, whether allelopathy or nutrient competition
195	played a causative role. Because R. ficaria has been identified as invasive (Axtell et al., 2010)
196	and has been associated with reduced native abundance and diversity (Hohman, 2005), we
197	expected that the presence of <i>R. ficaria</i> would have a negative impact on the performance of <i>I</i> .
198	capensis. Our results show that R. ficaria does indeed negatively affect I. capensis providing
199	confirmatory evidence of its assumed impact on native species. Ranunculus ficaria showed a
200	tendency to have a negative overall impact on the life span of <i>I. capensis</i> with plants dying on
201	average ~10 days earlier in the presence of R. ficaria. In the absence of carbon, R. ficaria
202	decreased seed production by ~50%, in part most likely through its effects on life span. As
203	expected, nutrients showed a significant effect on growth (in terms of height and stem diameter)
204	and seed production. If nutrient competition were a key factor in inhibition of <i>I. capensis</i> by <i>R</i> .

ficaria, we would have expected *I. capensis* to exhibit a release from competition in the presence of added nutrients. There was no significant interaction between nutrient addition and presence of *R. ficaria*, suggesting that nutrient competition was most likely not the primary mechanism of impact of *R. ficaria*. We do however acknowledge that our study does not rule out other forms of resource competition, such as competition for light or water.

210 In the presence of carbon, there was no effect of *R. ficaria*; carbon therefore ameliorated the 211 negative effect of *R. ficaria* on *I. capensis*. In previous research, application of carbon to soils 212 has mitigated the effects of invasive species (Ridenour and Callaway, 2001; Cipollini et al., 213 2008a). Ridenour and Callaway (2001) found that the competitive ability of an invasive 214 *Centaurea* species was greatly reduced by activated carbon and suggested that the advantage of 215 this species, at least in part, was due to allelopathy. Similarly, one possible conclusion from our 216 study is that the addition of carbon may have attenuated the allelopathic effect of *R. ficaria*. 217 However, when using activated carbon as an experimental tool to test allelopathy, caution must 218 be used in interpreting results. It has been shown that addition of activated carbon can change 219 soil conditions in potting soil (Lau et al., 2008; Weisshuhn and Prati, 2009), though it is 220 important to note that these studies used twice as much activated carbon compared to the amount 221 used in our study. Activated carbon can also affect root mutualisms, perhaps by absorbing 222 signaling compounds (Wurst *et al.*, 2010). Activated carbon itself can have a direct fertilizing effect (Lau et al., 2008). In our study, the mitigating effect of carbon was significant across both 223 224 nutrient treatments, suggesting that direct addition of nutrients by the activated carbon was most 225 likely not the mechanism through which carbon exerted its effects. *Ranunculus ficaria* contains a 226 number of secondary chemicals (Texier et al., 1984; Tomczyk et al., 2002) and is purported to 227 have medicinal uses in herbal medicine as a treatment for hemorrhoids and as an astringent

228 (Chevallier, 1996). The presence of putative bio-active secondary compounds may be a good 229 predictor of invasive species impacts, including allelopathy (Ehrenfeld, 2006). Further, because 230 *R. ficaria* exhibits direct allelopathy on some species in the laboratory and greenhouse (Cipollini, 231 unpublished data), allelopathy is a likely mechanism in the field, though clearly further study is 232 needed (see Blair et al., 2009). Whatever the exact mechanism of its effect, activated carbon 233 nevertheless was clearly able to negate the negative effect of *R. ficaria*, suggesting that, at the 234 very least, it may serve as an effective restoration tool to modify soil conditions in the field to the 235 benefit of native species (Kulmataski and Beard, 2006; Kulmatiski, 2010), provided it can be 236 effectively applied in restoration practice.

237 Interestingly, we found that *R. ficaria* had lasting effects beyond its brief growing season. 238 Hohman (2005) similarly found reduced diversity associated with presence of *R. ficaria* for 239 species other than ephemeral species, though the results were correlational rather than 240 experimental in nature. We expected that effects on *I. capensis* would not be particularly strong, 241 as it is thought that the negative effects of this species are exerted primarily on spring ephemeral 242 species (Swearingen, 2005). Further studies using spring ephemeral species are obviously needed 243 to clarify the comparative effect on this set of species presumed most sensitive. Even though R. 244 *ficaria* had completely senesced by week 6 of the experiment, it still significantly negatively 245 impacted I. capensis, which lived without the presence of R. ficaria for about two-thirds of its 246 life span. Other invasive species can have residual effects on native species, even after the 247 removal of the invasive species (Conser and Conner, 2009). The effect of *R. ficaria* well past its 248 growing season may be due to lingering effects, such as those due to allelochemicals or to other 249 modification of soil conditions. An alternative explanation may be that the seedling stage is the 250 most vulnerable stage of *I. capensis*, similar to the findings in Barto *et al.* (2010).

251 This is the first study to show the negative impact of *R. ficaria* on *I. capensis* and to point 252 towards a possible mechanism of success – allelopathy or other modification of soil conditions. 253 It is surprising that this is the first published study to investigate the invasive potential of R. 254 *ficaria*, given that natural resource agencies have recognized it as a species important to control 255 in natural areas since at least the year 2000 (Hammerschlag et al., 2002). Ranunculus ficaria has 256 already been banned in two states (Axtell et al., 2010) since the year 2006. Admittedly, there is a 257 balance between taking immediate action against an invasive species and waiting for time-258 consuming scientific studies (Blossey, 1999). Indeed, we support the use of the "precautionary" 259 principle" (e.g., Blossey et al., 2001) in assessing invasive species. Since R. ficaria can form 260 dense monocultures, the assumption that it is an invasive species is most likely a valid one. 261 However, in the ten or more years it has been identified as a concern, there is not a single 262 published paper documenting its impact. Even with our evidence of a negative impact of R. 263 ficaria, it is still entirely possible that the impact of R. ficaria has been exaggerated (Lavoie, 264 2010), causing larger-than-needed economic losses to the horticultural industry and redirection 265 of resources away from more important invaders. This research provides an example of the need 266 for more basic research into invasive species impacts and mechanisms of impacts on native 267 species for use in effective invasive species targeting, control and management.

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Source	df	F	Р
Presence of Ranunculus ficaria	1	0.88	0.352
Presence of Carbon	1	3.07	0.086
Nutrient Addition	1	45.30	<0.001
<i>R. ficaria</i> x Carbon	1	6.81	0.012
R. ficaria x Nutrients	1	0.05	0.828
Carbon x Nutrients	1	0.65	0.423
R. ficaria x Carbon x Nutrients	1	0.96	0.331
Error	47		

TABLE 1.-Results of ANOVA on total seed number of Impatiens capensis

Source	df	Wilk's λ	F	Р
Starting Height Covariate	2	0.6732	5.31	0.013
Presence of Ranunculus ficaria	2	0.9004	1.216	0.315
Presence of Carbon	2	0.9075	1.121	0.344
Nutrient Addition	2	0.4247	14.904	< 0.001
<i>R. ficaria</i> x Carbon	2	0.8954	1.286	0.296
R. ficaria x Nutrients	2	0.8838	1.447	0.257
Carbon x Nutrients	2	0.9455	0.635	0.540
R. ficaria x Carbon x Nutrients	2	0.7831	3.047	0.068
Error	22			

TABLE 2.-Results of MANCOVA on height and stem diameter of Impatiens capensis on August

14 (week 16)

Source	df	F	Р
Starting Height Covariate	1	9.46	0.005
Presence of Ranunculus ficaria	1	1.19	0.287
Presence of Carbon	1	0.05	0.831
Nutrient Addition	1	7.28	0.013
R. ficaria x Carbon	1	0.31	0.584
R. ficaria x Nutrients	1	2.19	0.153
Carbon x Nutrients	1	1.33	0.261
R. ficaria x Carbon x Nutrients	1	0.58	0.454
Error	23		

TABLE 3.-Results of ANCOVA on height of Impatiens capensis on August 14 (week 16)

Source	df	F	Р
Starting Height Covariate	1	8.59	0.008
Presence of Ranunculus ficaria	1	2.53	0.126
Presence of Carbon	1	1.12	0.302
Nutrient Addition	1	30.34	< 0.001
<i>R. ficaria</i> x Carbon	1	0.75	0.395
R. ficaria x Nutrients	1	0.04	0.896
Carbon x Nutrients	1	0.52	0.479
R. ficaria x Carbon x Nutrients	1	2.01	0.170
Error	23		

TABLE 4.-Results of ANCOVA on stem diameter of Impatiens capensis on August 14 (week 16)

Figure captions

FIG. 1—Mean (\pm SE) life span of *Impatiens capensis* grown with and without *Ranunculus ficaria*, across nutrient and carbon treatments

FIG. 2—Mean (\pm SE) number of seeds, height and stem diameter of *Impatiens capensis* grown with and without nutrients, across *Ranunculus ficaria* and carbon treatments.

FIG. 3—Mean (\pm SE) number of seeds for *Impatiens capensis* grown with and without *Ranunuculus ficaria* in the presence and absence of carbon, across nutrient treatments





